

Charcot-Marie-Tooth disease: correction of the CMT-1A phenotype

Shelley L. Davies, M^a Angels Moral

Prous Science, P.O. Box 540, 08080 Barcelona, Spain

CONTENTS

Abstract	531
Introduction	531
Targets and therapeutic advances	531
References	532

Abstract

Charcot-Marie-Tooth (CMT) disease constitutes a group of prevalent hereditary, chronic and debilitating peripheral neuropathies. CMT type 1A (CMT-1A), the most common form of CMT, is autosomal dominant and is characterized by peripheral demyelination. No pharmacotherapies currently exist for CMT-1A, although identification of an underlying duplication in the gene for PMP22 (peripheral myelin protein 22), a glycoprotein expressed in myelin, has ignited the search for candidates to correct the CMT-1A phenotype. To date, these include the progesterone antagonist onapristone, the antioxidant ascorbic acid and the natural neurotrophic factor neurotrophin-3.

Introduction

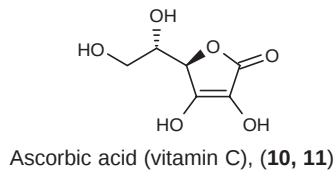
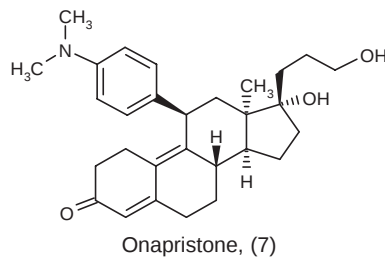
Charcot-Marie-Tooth (CMT) disease, named after the three physicians who first reported it in 1886, but also sometimes known as hereditary motor and sensory neuropathy (HMSN) or peroneal muscular atrophy, constitutes a group of genetic peripheral neuropathies that cause muscle weakness and wasting in the feet, legs, hands and forearms, as well as sensory loss in the limbs. CMT disease is estimated to affect 1 in 2,500 individuals in the U.S. alone and is the most common inherited neurological disorder. Autosomal dominant, autosomal recessive and X-linked forms have been recognized. CMT type 1A (CMT-1A), the most common form of CMT, is autosomal dominant and is characterized by peripheral demyelination and slow nerve conduction (< 38 m/s). At present, there is no pharmacotherapy for CMT-1A and current treatment options are aimed at helping patients cope with the disabling symptoms of the disease, via physical or occupational therapy, orthopedic devices and orthopedic surgery.

Targets and therapeutic advances

CMT-1A involves a 1.5-Mb duplication of the p11.2-p12 short-arm region of chromosome 17 (1, 2), with gene mapping indicating that the gene for PMP22 (peripheral myelin protein 22), a glycoprotein expressed in myelin, is located within this region (3). The development of animal models to mimic the CMT-1A phenotype, via *PMP22* gene overexpression and point mutations, has indicated that the severity of CMT-1A is dependent on the level of PMP22 overexpression (4). Therefore, PMP22 overexpression is an important target for novel CMT-1A treatment options, and several recent studies have identified potential candidates (outlined in Figure 1) to correct the CMT-1A phenotype.

Studies have suggested that the steroid hormone progesterone, an epigenetic regulator of gene expression, can stimulate PMP22 expression (5, 6). In accordance, daily administration of the progesterone antagonist onapristone (Schering AG) at a dose of 20 mg/kg over a period of 7 weeks has been shown to induce a 15% decrease in PMP22 mRNA in sciatic nerves in a rat model of CMT-1A. This was associated with a 1.5-fold increase in *Mpz* mRNA, which encodes the major structural protein of myelin. Onapristone-treated rats also maintained a significantly greater number of sciatic nerve axons and exhibited improved motor performance after 5 weeks of treatment (+65%) (7).

Further studies tested the promyelination properties of the antioxidant ascorbic acid (vitamin C) (8, 9) in CMT-1A mice. Weekly administration of ascorbic acid (1.12 mg) to male CMT-1A mice produced a significant decrease in locomotor deterioration within 1 month of treatment, with further analyses confirming an associated recovery of muscle strength (via grip tests). Treatment continuation also provided a significant improvement in mean lifespan (19.7 months vs. 6 months for untreated mice), which was comparable to the average lifespan of normal mice. Histological analysis of CMT-1A peripheral nerves indicated that these improvements in behavior were associated with an increased number of myelinated axons (70% on ascorbic acid vs. 25% on placebo) and a 10-fold lower level of PMP22 expression (10). A phase II clinical study to assess the efficacy and safety of ascorbic acid in



```
MSILFYVIFLAYLRGIQGNMDQRLPEDSLNSLIKLIQADILK
NKLSKQMVVKENYQSTLPKAEAPREPERGGPAKSAFQPIAMDT
ELLRQQRRYNSPRVLLSDSTPLEPPPLYLMEDYVGSPVVANRTSR
RKRYAEHKSHRGEYSVCDSESLWVTDKSSAIDIRGHQVTVLGEIK
TGNSPVKQYFYETRCKEARPVKNGCRGIDDKHWN SQCKTSQTYVR
ALTSENNKLVGWRWIRIDTSCVCALSRKIGRT
```

Neurotrophin-3, (12)

Fig. 1. Therapeutic candidates for CMT-1A.

young CMT-1A patients is expected to begin shortly at the University of Amsterdam (11).

Neurotrophic agents may also be useful to promote axonal regeneration and myelination in CMT-1A. Application of neurotrophin-3 (NT-3; 5 mg/kg s.c. 3x week), a crucial Schwann cell component involved in survival and differentiation, was investigated in two CMT-1A mouse models: 1) nude mice harboring CMT-1A xenografts with characteristic delayed myelination; and 2) Trembler^J mice, which carry a point mutation in the *PMP22* gene and demonstrate characteristic impairment of nerve regeneration following crush injury. In both models, NT-3 significantly enhanced myelinated fiber densities, with further evidence to suggest normalization of Schwann cell-axonal interactions in CMT-1A xenografts and significantly improved Schwann cell density in the sciatic nerves of Trembler^J mice. Based on its *in vivo* efficacy, NT-3 was assessed in a clinical trial in CMT-1A patients. A double-blind, placebo-controlled, randomized, pilot study in 8 patients receiving human recombinant NT-3 (150 mg/kg 3x week over 24 weeks) indicated that NT-3 treatment significantly improved sensory and reflex scores, but did not affect motor scores. Examination of sural nerve biopsies from these patients indicated an associated augmentation of myelinated fiber and Schwann cell regeneration units (12).

References

1. Lupski, J.R., Oca-Luna, R.M., Slaugenhaupt, S. et al. *DNA duplication associated with Charcot-Marie-Tooth disease type 1A*. Cell 1991, 66(2): 219-32.
2. Raeymaekers, P., Timmerman, V., Nelis, E. et al. *Duplication in chromosome 17p11.2 in Charcot-Marie-Tooth neuropathy type 1a (CMT 1a)*. The HMSN Collaborative Research Group. Neuromusc Disord 1991, 1(2): 93-7.
3. Matsunami, N., Smith, B., Ballard, L. et al. *Peripheral myelin protein-22 gene maps in the duplication in chromosome 17p11.2 associated with Charcot-Marie-Tooth 1A*. Nat Genet 1992, 1(3): 176-9.
4. Huxley, C., Passage, E., Robertson, A.M. et al. *Correlation between varying levels of PMP22 expression and the degree of demyelination and reduction in nerve conduction velocity in transgenic mice*. Hum Mol Genet 1998, 7(3): 449-58.
5. Melcangi, R.C., Ballabio, M., Cavarretta, I., Gonzalez, L.C., Leonelli, E., Veiga, S., Martini, L., Magnaghi, V. *Effects of neuroactive steroids on myelin of peripheral nervous system*. J Steroid Biochem Mol Biol 2003, 85(2-5): 323-7.
6. Schumacher, M., Guennoun, R., Mercier, G. et al. *Progesterone synthesis and myelin formation in peripheral nerves*. Brain Res Brain Res Rev 2001, 37(1-3): 343-59.

7. Sereda, M.W., Meyer zu Horste, G., Suter, U., Uzma, N., Nave, K.A. *Therapeutic administration of progesterone antagonist in a model of Charcot-Marie-Tooth disease (CMT-1A)*. Nat Med 2003, 9(12): 1533-7.
8. Eldridge, C F., Bunge, M.B., Bunge, R.P. *Differentiation of axon-related Schwann cells in vitro: II. Control of myelin formation by basal lamina*. J Neurosci 1989, 9(2): 625-38.
9. Podratz, J.L., Rodriguez, E.H., Windebank, A.J. *Antioxidants are necessary for myelination of dorsal root ganglion neurons, in vitro*. Glia 2004, 45(1): 54-8.
10. Passage, E., Norreel, J.C., Noack-Fraissignes, P. et al. *Ascorbic acid treatment corrects the phenotype of a mouse model of Charcot-Marie-Tooth disease*. Nat Med 2004, 10(4): 396-401.
11. *Ascorbic Acid Treatment in CMT1A Trial (AATIC)* (NCT00271635). ClinicalTrials.gov Web Site 2006, January 3.
12. Sahenk, Z., Nagaraja, H.N., McCracken, B.S., King, W.M., Freimer, M.L., Cedarbaum, J.M., Mendell, J.R. *NT-3 promotes nerve regeneration and sensory improvement in CMT1A mouse models and in patients*. Neurology 2005, 65(5): 681-9.